



UNIVERSITY OF CALGARY FACULTY OF VETERINARY MEDICINE

This review accompanies the relevant episode of the Cutting Edge veterinary podcast. In each episode of this podcast, 3rd year students in the University of Calgary's veterinary medicine program fill you in on the most up-to-date literature and evidence-based practices on topics that matter to you, the practising veterinarian.

A Comparison of Caseous Lymphadenitis (*Corynebacterium pseudotuberculosis*) Treatments In Small Ruminants

STUDENTS: Emma Mandolesi-Kahanyshyn, Sarah Kulle and Allison Kwantes

FACULTY MENTORS: Drs. Brielle Rosa and Karen Liljebjelke

Background

Caseous lymphadenitis (CLA) is a chronic, highly infectious disease of economic importance found worldwide in small ruminants, costing 15-17 million dollars annually^{1,2}. Economic consequences include carcass condemnation, decreased production, reproductive losses, and death^{1,3}. Historically, CLA was implicated as the third leading cause of economic loss to the sheep industry of the western United States⁴. CLA impacts both sheep and goats, however, despite goats having more CLA lesions at slaughter, research has indicated that there is higher prevalence and morbidity in sheep^{5,6}. A 2009 study surveying over one thousand small ruminants determined prevalence of CLA to be 50.47%⁵. In some countries, CLA is not a notifiable disease, therefore, it has been suggested that the true prevalence is currently underestimated⁷. This emphasizes the importance of CLA, and why it is essential to have effective treatments to enhance animal welfare and economics⁵. Canadian goat and sheep production has gradually increased in the past decade^{8,9}. Alberta is the second leading province in goat meat production and is ranked among the top three sheep producing provinces^{8,10}. The economic implications associated with CLA are even more relevant for Canada's small and growing sector, warranting a review of CLA management options to protect Canadian flocks.

The pathological agent, *Corynebacterium pseudotuberculosis*, is a gram-positive aerobe which causes specific disease syndromes in sheep, goats and horses^{1,11}. *C. pseudotuberculosis* targets the lymphatic system and can cause superficial and/or visceral lymphadenopathy, and in the skin and viscera it is commonly characterized by caseous abscessation. Pathogen spread can occur due to various management practices on farms (ie. shearing, tail docking), environmental contamination from lesions rupturing, or by direct contact between animals via skin penetration or respiratory/oral routes^{1,4,12}. Once an individual is infected, the disease is difficult to eliminate due to poor responses to therapeutics, pathogen virulence factors, and diagnostic limitations^{1,11,12}.

The most widely accepted control measures for CLA includes the treatment of valuable individuals, aggressive culling, implementation of strong biosecurity protocols, and/or

vaccination^{1,4}. In this review, we focus on individual animal treatments, specifically comparing open drainage with lavage and closed system lesion management with intralesional or systemic antimicrobial therapy. The proposed benefit of closed-system management is the reduced risk of environmental contamination. Surgical removal also reduces this risk; however, it has higher costs and increased risks associated with general anesthesia^{1,4}. Antimicrobials that have shown some efficacy against CLA include penicillin, oxytetracycline, and tulathromycin^{1,4,13}. We reviewed the literature to determine the susceptibility of *C. pseudotuberculosis* to the stated antimicrobials, as well as their efficacy in treating CLA. Additionally, we examine the efficacy of different vaccines in treating and/or preventing CLA in small ruminants.

Closed versus open lavage

As mentioned, one option for the individual CLA treatment is lavage, which can be done either via closed or open methods. The research in this area is limited not only by the number of publications but also the number of individuals included in each study. There is no clear evidence to say which method is most effective. A 2009 study is the most current research that comes closest to comparing these methods; however, their use of different antimicrobial drugs confounds the comparison of such lavage methods⁴. This study included three treatment groups: Group 1 was treated via open drainage, saline lavage and subcutaneous (SC) procaine penicillin G (PPG) injection, Group 2 was treated with closed lavage of lesions and an intralesional injection of tulathromycin, and Group 3 was treated with closed lavage and a SC injection of tulathromycin. From this study, the authors concluded that needle distension lavage combined with tulathromycin could be an acceptable alternative to open drainage and flushing. This was based on the similar resolution proportions of 92.9%, 83.3% and 82.4% for groups 1, 2 and 3, respectively. A suggested benefit is a decrease in environmental contamination when using closed system drainage. Note that the study lacked a control group comparing whether closed system lavage without an antibiotic was comparable.

Another study compared two intralesional injection groups and a control group that used the conventional treatment method of open drainage and iodine lavage¹¹. Iodine tincture and sodium hypochlorite were used as intralesional treatments and although using intralesional injections is not the focus of this review, the results could be used to consider the implications of closed drainage and intralesional injection of antimicrobials. In both intralesional groups, natural rupture occurred in all animals' post-treatment. The material from five out of the six ruptured lesions tested positive for *C. pseudotuberculosis*, therefore failing to reduce the risk of environmental contamination. Edema was also found in the regions of the lymph nodes belonging to the intralesional treatment groups¹¹. This was proposed to be due to irritation of the products used at high concentrations. One advantage suggested from using intralesional treatments was a shortened clinical stage of disease (which in this study was one month shorter than the conventional treatment). Like the study described above, due to the small sample size, incompletely described methods and an unclear control group, there is weak evidence to make conclusive statements regarding closed versus open drainage.

Antimicrobials

Antimicrobials are commonly used to treat infections in small ruminants but are often used in an extra-label manner since there are few drugs approved for small ruminants in

Canada. With regards to CLA, there are data available on outcomes after treatment with oxytetracycline, penicillin and tulathromycin, with penicillin and oxytetracycline as the most used treatments in practice¹⁴. The use of tulathromycin has been explored due to its ability to penetrate tissue chambers in bovine and swine, suggesting a potential new solution to CLA management¹⁵.

Oxytetracycline:

Oxytetracycline (OTC) is a broad-spectrum antimicrobial drug which distributes to most tissues; It has historically been used in the treatment of gastrointestinal and respiratory disease^{13,16}. It is effective against aerobic gram-positive and gram-negative bacteria and has been suggested to be effective against *C. pseudotuberculosis*^{13,17}. Reported doses of injectable OTC range from 2-20 mg/kg SID or q3-5d depending on the formulation used^{13,17,18}. OTC may be effective at penetrating encapsulated lesions due to its moderate lipid solubility¹³. Specifically, in comparison to tulathromycin, suggested benefits of OTC include lower cost, higher availability, and a shorter withdrawal time. Currently in the literature there is only one study with weak to moderate evidence supporting the use of OTC as a treatment for CLA¹³. Treatment groups consisted of sheep that received an OTC at a dose of 20 mg/kg administered by percutaneous injection into a chamber inoculated with *C. pseudotuberculosis*, percutaneous injection into an uninoculated control chamber, or IM injection in the cervical region. Results indicated sheep that received OTC via IM injection achieved drug concentrations >1 ug/mL (MIC for isolate, 0.5 ug/mL) in all *C. pseudotuberculosis* inoculated tissue chambers but these all remained culture positive for *C. pseudotuberculosis* throughout the entire study. The authors suggested that MIC levels were not maintained long enough to eliminate *C. pseudotuberculosis*. In comparison, intra-chamber administration of OTC effectively eliminated the *C. pseudotuberculosis* isolate <48 hours after treatment. *In situ* efficacy would require further studies to determine.

Penicillin:

There are limited data on the pharmacokinetics and pharmacodynamics of penicillin in goats. Doses for goats are often extrapolated from sheep or cattle doses, despite the differences in metabolism that exist between these species^{19,20}. For small ruminants, penicillin has historically been used for the treatment of clinical mastitis, diarrhea and pneumonia¹⁴. It has also become an option for CLA treatment in part since it has a broad spectrum of action and covers multiple gram-positive organisms, therefore covering co-infections that may arise⁴. PPG has been approved for used in sheep in USA, (with the dose of 660U/kg of body weight for 4-7days) but still is not approved for goats¹⁹. Penicillin is the most common drug residue detected in tissue and milk, and although rare, anaphylactic reactions resulting from meat residues have been noted^{19,20}. Due to this, and the fact that penicillin is off label for goats, there can be no detectable levels of penicillin in tissues which complicates penicillin at a CLA treatment choice. Derbyshire's study of mastitis in goats indicated that goats with mild to moderate mastitis infections responded well to penicillin, but those with severe infections were unable to recover, which begs the question how effective penicillin may be for severe CLA cases, especially with extensive visceral involvement²¹. On the other hand, in the 2009 study by Washburn, intralesional PPG injection (20,000U/kg) yielded better results than intralesional tulathromycin injection⁴. Additionally, Al-Gaabary et al's study examining sheep treated with PPG on shearing days showed that with sufficient hygienic measures, prophylactic PPG injections were effective

at minimizing CLA prevalence in affected flocks⁵. Although there is little evidence supporting its use as a CLA treatment, penicillin may have some efficacy for CLA treatment and prevention.

Tulathromycin:

The pharmacological properties of tulathromycin may justify its use in the treatment of CLA in sheep and goats. Tulathromycin has a high volume of distribution and is eliminated slowly; providing a prolonged period of tissue concentration above the MIC for common bacterial pathogens of the bovine and swine respiratory tract^{15,22}. Tulathromycin also has gram positive activity; therefore, it should be effective against *C. pseudotuberculosis*^{15,22}. The safety of tulathromycin use in goats has been demonstrated in small-scale studies involving young juveniles and mature adults^{23,24}. Washburn et al. demonstrated that 25 mg/kg SC injections (ten times the recommended dose in cattle and swine) were well tolerated in yearling goats, aside from a transiently increased creatine kinase and some genotoxic effects²³. In a different study, 2.5, 7.5, and 12.5 mg/kg SC injections given weekly for three weeks were all well tolerated in juvenile goats, with no significant differences in physical or histological parameters²⁴. The ability to safely give multiple injections is considerable due to the antimicrobial's primarily bacteriostatic mechanism of action^{15,22}. The known safety of repeat injections would allow practitioners to treat persistent abscesses with more confidence. As for efficacy of tulathromycin in the closed-system treatment of CLA, the evidence is not strong. Neither the use of single dose (2.5 mg/kg) intralesional or SC tulathromycin combined with closed system lavage was significantly better in resolving superficial CLA compared to open-system lavage and SC PPG⁴. Intralesional injection of 2.5 mg/kg tulathromycin into implanted tissue chambers did maintain a concentration above the pathogen's MIC (2 ug/mL) for 15 days; however, some chambers showed no decline in pathogen growth²⁸. Additionally, intralesional injections had the same success rate as SC injections for inhibiting pathogen growth; suggesting that both routes of administration could be acceptable²⁸. Overall, tulathromycin has been shown to be safe to use extra-label in goats, however it's efficacy in treating CLA is mixed. Further studies are required to determine true efficacy, as well as studies from the field to determine efficacy *in situ*.

Within the considerations for any antibiotic used, the importance of antimicrobial resistance (AMR) cannot be overstated. Since there is little on-label data for small ruminants, if not given proper protocols, producers may overestimate body weight and often give dosages higher than the ones prescribed¹⁴. In a 2017 study, milk from 100% of the goat bulk tanks (n = 26) and 87% of the sheep bulk tanks (n =47) sampled contained bacteria with resistance to penicillin, emphasizing the potential for the development of AMR in these species³⁰. Additionally, we must consider the importance of these antimicrobials to human medicine. Oxytetracycline and penicillin are highly important antimicrobial drugs, whereas tulathromycin is a critically important antimicrobial according to the World Health Organization³¹. These classifications further emphasize the importance of antimicrobial stewardship if practitioners do choose to use these antimicrobials.

Vaccines

In addition to using lavage or antimicrobials, vaccine treatments have also been attempted to treat or prevent CLA. Vaccines have the potential to be an effective preventative measure or treatment during a CLA outbreak, especially due to their ease of delivery and safety³². Currently, the most accepted vaccine for treating CLA is Glanvac-6, but Case-Bac is also

available in Canada^{6,33}. Both are toxoid vaccines labelled only for sheep; there are no vaccines labelled for use in goats in Canada for the management of CLA^{12,33,34}. Attempts have been made to use other formulations (live, DNA); however, these had limited ability to treat CLA. Glanvac-6 has been shown to limit the spread of infection beyond the initial infection site but provides varied protection, ranging from 25-90% reduction in new CLA lesions following vaccination of a previously unvaccinated herd^{6,32}.

In terms of using vaccines as a protective measure, there have been limited studies indicating their functionality. In a Sutherland et al study 81 sheep were challenged with *C. pseudotuberculosis* three months after completing a two-vaccination series against CLA, with 27 sheep still developing CLA lesions². This could indicate that vaccines may not effectively prevent CLA, but more research is required to deepen understanding. In addition to the Sutherland et al study, Windsor suggested that vaccines may not be the most effective method of CLA prevention since *C. pseudotuberculosis* shedding may still occur years after vaccination³². Although data are limited on the benefits of prophylactic preventative vaccines, multiple sources have determined that a protocol of at least two doses administered one month apart, in addition to annual boosters, (and according to Windsor, potentially more vaccinations around lambing and weaning season), are needed to have any successful prophylactic protection^{32,35}. Ultimately, vaccinations can decrease microbial contamination, therefore reducing spread of disease and associated economic and animal losses; but completely eliminating *C. pseudotuberculosis* infections is unlikely.

Acknowledgments

We would like to thank Dr. Brielle Rosa and Dr. Karen Liljebjelke for their assistance and mentorship in conducting this review and writing this manuscript.

References:

1. Williamson LH. Caseous lymphadenitis in small ruminants. *Vet Clin North Am Food Anim Pract.* 2001;17(20):359-371.
2. Sutherland SS, Ellis TM, Paton MJ, Mercy AR. Serological response of vaccinated sheep after challenge with *Corynebacterium pseudotuberculosis*. *Aust Vet J.* 1992;69(7):168–169.
3. Santos LM, Stanisic D, Menezes UJ, Mendonça MA, Barral TD, Seyffert N, et al. Biogenic silver nanoparticles as a post-surgical treatment for *Corynebacterium pseudotuberculosis* infection in small ruminants. *Front Microbiol.* 2019;10:1–11.
4. Washburn KE, Bissett WT, Fajt VR, Libal MC, Fosgate GT, Miga JA, et al. Comparison of three treatment regimens for sheep and goats with caseous lymphadenitis. *J Am Vet Med Assoc.* 2009;234(9):1162–6.
5. Al-Gaabary MH, Osman SA, Oreiby AF. Caseous lymphadenitis in sheep and goats: Clinical, epidemiological and preventive studies. *Small Rumin Res.* 2009;87(1-3):116–21.
6. Windsor PA. Control of Caseous Lymphadenitis. *Vet Clin North Am Food Anim Pract.* 2011;27(1):193–202.
7. Abebe D, Tessema S. Determinization of *Corynebacterium pseudotuberculosis* prevalence and antimicrobial susceptibility pattern of isolates from lymph nodes of sheep and goats at an organic export abattoir, Modjo, Ethiopia. *Lett Appl Microbiol.* 2015;61(5):469-476.
8. Alberta Agriculture, Food and Rural Development. Agriculture business profiles: commercial meat goat industry. Alberta: Alberta Agriculture, Food and Rural Development; 2006. 9 p. Agdex. No.: 435/830-1.
9. Lu CD, Miller BA. Current status, challenges and prospects for dairy goat production in the Americas. *Asian-Australas J Anim Sci.* 2019; 32(8):1244-1255.
10. Agriculture and Agri-Food Canada. Sheep and lamb quick facts 2021. Ontario: Agriculture and Agri-Food Canada; 2021 [Cited 2023 Feb 17]. Available from: <https://agriculture.canada.ca/en/sector/animal-industry/red-meat-and-livestock-market-information/sheep-and-lamb>
11. Santiago LB, Pinheiro RR, Alves FSF, Santos VWS, Rodrigues AS, Lima AMC, et al. *In vivo* evaluation of antiseptics and disinfectants on control of Caseous Lymphadenitis: clinical, hematological, serological and microbiological monitoring. *Arq Inst Biol.* 2013;80(3):273-280.
12. Smith MC, Sherman DM. Subcutaneous Swellings. In: *Goat Medicine.* John Wiley & Sons, Incorporated; 2022. p. 67–94.
13. Washburn KE, Fajt VR, Polasek AN, Lawhon SD, Padgett AL, Lo CP, et al. Oxytetracycline concentrations in interstitial fluid from tissue chambers inoculated with *Corynebacterium pseudotuberculosis* after intramuscular or intrachamber administration in sheep. *Am J Vet Res.* 2019;80(60):586-594.
14. Lianou DT, Fthenakis GC. Use of antibiotics against bacterial infections on dairy sheep and goat farms: patterns of usage and associations with health management and human resources. *Antibiotics.* 2022;11(6):753-774.
15. Evans NA. Tulathromycin: an overview of a new triamilide antibiotic for livestock respiratory disease. *Vet Ther.* 2005 Summer; 6(2):83-95.
16. Tardiveau J, LeRoux-Pullen L, Gehring R, Touchais G, Chotard-Soutif MP, Mirfendereski H et al. A physiologically based pharmacokinetic (PBPK) model exploring the blood-milk barrier in

- lactating species- A case study with oxytetracycline administered to dairy cows and goats. *Food Chem Toxicol.* 2022;161.
17. Aktas I, Yarsan E. Pharmacokinetics of Conventional and Long-Acting Oxytetracycline Preparations in Kilis Goat. *Front Vet Sci.* 2017;4(229).
 18. Yucel UM, Kosal V, Kara M, Taspinar F, Uslu BA. Adverse effects of oxytetracycline and enrofloxacin on the fertility of Saanen bucks. *Trop Anim Health Prod.* 2021;53(466):1-7.
 19. Li M, Gehring R, Riviere JE, Lin Z. Development and application of a population physiologically based pharmacokinetic model for penicillin G in swine and cattle for food safety assessment. *Food Chem Toxicol.* 2017;107:74–87.
 20. Payne MA, Craigmill A, Riviere JE, Webb AI. Extralabel use of penicillin in Food Animals. *J Am Vet Med Assoc.* 2006;229(9):1401–1403.
 21. Derbyshire JB. Treatment of experimental streptococcal and staphylococcal mastitis in the goat with penicillin. *Journal of Comparative Pathology and Therapeutics.* 1964;74:31–36.
 22. Draxxin. In: *Compendium of Veterinary Products [on-line]*. Kirkland (QC): Zoetis Canada. [February 17, 2023]. Available from: vetalytixcanada.cvp-service.com
 23. Washburn KE, Bissett W, Fajt V, Clubb F, Fosgate GT, Libal M, Smyre KE, Cass KL. The safety of tulathromycin administration in goats. *J Vet Pharmacol Therap.* 2007; 30:267-270.
 24. Clothier KA, Jordan DM, Loynachan AT, Griffith RW. Safety evaluation of tulathromycin use in the caprine species: tulathromycin toxicity assessment in goats. *J Vet Pharmacol Therap.* 2010;33:499-502.
 25. Martin KL, Clapham MO, Davis JL, Baynes RE, Lin Z, Vickroy TW, Riviere JE, Tell LA. Extralabel drug use in small ruminants. *J Am Vet Med Assoc.* 2018;253(8):1001-1009
 26. Young G, Smith GW, Leavens TL, Wetzlich SE, Baynes RE, Mason SE, Riviere JE, Tell LA. *Res Vet Sci.* 2011; 90:477-479.
 27. Amer AMM, Constable PD, Goudah A, El Badawy SA. Pharmacokinetics of tulathromycin in lactating goats. *Small Rumin Res.* 2012;108:137-43.
 28. Washburn KE, Fajt VR, Lawhon SD, Adams LG, Tell LA, Bissett WT. Caprine abscess model of tulathromycin concentrations in interstitial fluid from tissue chambers inoculated with *Corynebacterium pseudotuberculosis* following subcutaneous or intrachamber administration. *Antimicrob Agents Chemother.* 2013;57(12):6295-6304.
 29. Santman-Berends I, Luttikholt S, Brom RV, Schaik GV, Gonggrijp M, Hage H, et al. Estimation of the use of antibiotics in the small ruminant industry in the Netherlands in 2011 and 2012. *PLoS ONE.* 2014;9(8):1–10.
 30. Obaidat MM, Bani Salman AE, Roess AA. High prevalence and antimicrobial resistance of *Meca Staphylococcus aureus* in dairy cattle, sheep, and goat bulk tank milk in Jordan. *Trop Anim Health Prod.* 2017;50(2):405–12
 31. World Health Organization. Critically important antimicrobials for human medicine, 6th revision. Geneva: World Health Organization; 2019. 52 p.
 32. Windsor P. Managing control programs for ovine caseous lymphadenitis and paratuberculosis in Australia, and the need for persistent vaccination. *Veterinary Medicine: Research and Reports.* 2014;11-22.
 33. de Pinho RB, de Oliveira Silva MT, Bezerra FSB, Borsuk S. Vaccines for caseous lymphadenitis: up-to-date and forward-looking strategies. *App Microbiol Biotech.* 2021;105:2287-2296.

34. Inspection Canada. Biosecurity planning guide for Canadian goat producers [Internet]. Ontario: Inspection Canada. 2013 Aug 01 [Cited 2023 Feb 17]. Available from: <https://inspection.canada.ca/animal-health/terrestrial-animals/biosecurity/standards-and-principles/producer-guide-goats/eng/1375213342187/1375213659306?chap=0>
35. Stanford K, Brogden KA, McClelland LA, Kozub GC, Audibert F. The incidence of caseous lymphadenitis in Alberta sheep and assessment of impact by vaccination with commercial and experimental vaccines. *Can J Vet Res.* 1998;62(1):38-43